## Progress Towards an Intervention to Prevent Transfusion-Transmitted Babesia

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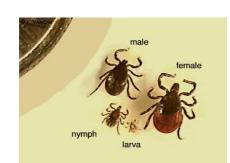
Transmissible Diseases Department American Red Cross Holland Laboratory *and* Department of Microbiology and Tropical Medicine George Washington University

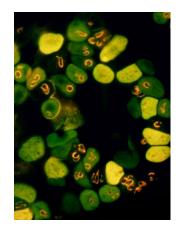
**Holland Laboratory** 

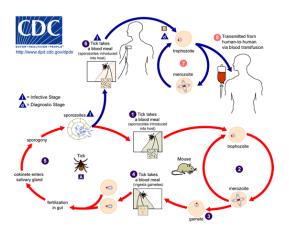


## Babesia spp.

- agents of human babesiosis:
  - B. microti, B. divergens & B. duncani
  - CA-1, MO-1, EU-1, KO-1, TW-1, etc.
- infect red blood cells, but occasionally found extracellular
- transmitted by *Ixodes* ticks (aka, the deer tick)
  - often same species that locally transmits Lyme borreliosis
- generally causes benign flu/malaria-like illness
- but can be fatal in:
  - infants
  - elderly
  - immunocompromised
    - sickle cell disease
  - asplenic









## **B. microti: Survival In Blood Products**



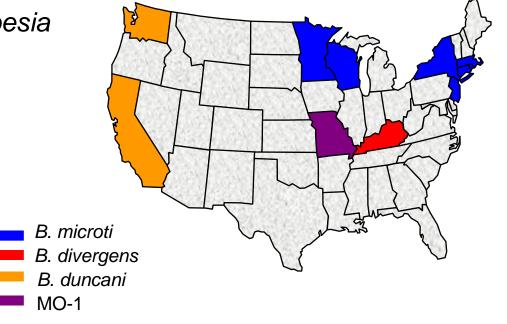
survives in red cells maintained at 4°C

- 21 days experimentally
- 42 days in association with transfusion case
- survives indefinitely in cryopreserved red cells
- parasite killed in frozen plasma
- extracellular parasites reported
  - pose potential issues for platelet apheresis & fresh plasma products



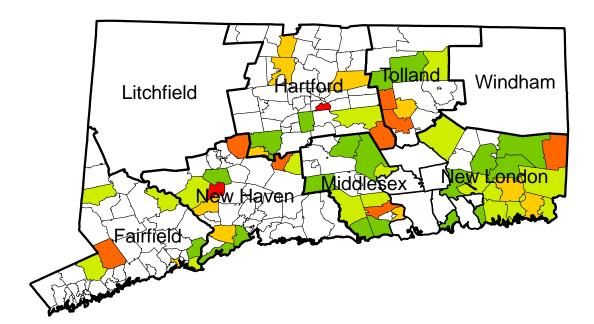
#### Babesia in the U.S.

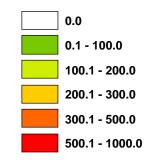
- 1993 B. duncani on West Coast
- 1996 MO1 in Missouri
- 1999 B. microti reported in New Jersey
- 2002 B. divergens in Kentucky
- other miscellaneous Babesia





## **Seroprevalence in Connecticut**







Spatial cluster 1

Spatial cluster 2

Johnson et al., Transfusion 2009;49:2574-2582



# **Seroprevalence in WI and MN**

- testing 2000 samples
  - initiated in October 2010
- focused on high case prevalence counties/cities
  - based on MN Health Department data
- all samples tested by IFA
  - positive samples tested by PCR
  - no opt-out option
- tested 574 samples to date
  - 5 (0.9%) IFA positive donors



#### Summary of 10 NCBS Transfusion Transmitted (TT) Babesia Investigations Since 7/2008

Case #	NCBS IFA	Product	# of Patients	Comments
	🕀 Donor	Involved	Infected	
1				Donor confirmed as source of
	No	RBC	0	Anaplasma infection. Negative for Babesia
2	Yes	Double RBC	2	1 Fatality
3	Yes	RBC	1	
4	Yes	RBC	3	Decedent and 2 Kidney txp recipients
5	Yes	RBC	1	
6	Yes	RBC	1	
$7^{*\dagger}$	No	RBC	1	Non ARC donor was ⊕
8 <sup>‡</sup>				Out of Region PRTTI.
	No	RBC	1	Another ARC region had + donor
9	Yes	RBC	1	
10	Yes	RBC	1	
	7* 7	8†	12‡	
Tetel	⊕ Donors	NCBS	Patients	
Totals		Implicated	(11 from MN	
		RBCS	Area)	

\*7NCBS 🕀 Donors but 8 MN donors as one donor was Non ARC See Case #7 above.

†8 NCBS RBCs but 9 MN RBCS because of Case #7 above .

‡ Case #8 above: Suspected NCBS RBC was exported to another ARC Region. ⊕ Patient &Donor not from MN.

#### **9** Potential Cases of Transfusion-Transmitted Babesia

- 11 Local Patients Affected
- 8 Local Donors Implicated (1 Case Non ARC)



#### > 100 cases associated with *B. microti*

3 cases associated with *B. duncani* 0 cases associated with other species, types, strains, etc.



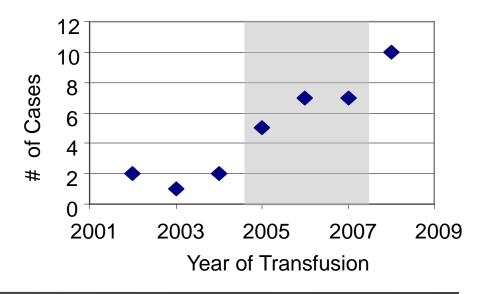
#### **B. microti: Transfusion Cases**

- > 100 known cases worldwide (1979 present)
  - 1 in Japan (autochthonous)
  - 1 in Canada (U.S. derived)
  - rest in U.S.
  - ~ 10 per year
- one possible case in Europe
  - Hildebrandt et al., Eur J Clin Microbiol Infect Dis 2007;26:595-601
- recipients neonates to 79 years
- fatalities increasingly reported
- red cells and whole blood platelets implicated
- no licensed tests
- gaining traction as critical blood safety issue



#### ARC Hemovigilance: 2005-2007

- suspected transfusion-transmitted *B. microti* infections reported by transfusion services
- additional cases through recipient tracing
- donor follow-up samples tested by IFA and PCR
- 19 cases transfusion-transmitted B. microti
  - 5 fatalities
  - 18 RBC units (1 split unit)



Tonnetti et al., Transfusion 2009;49:2557-2563

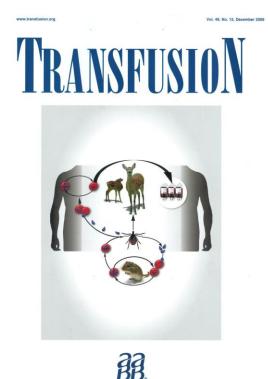
- 13 (68%) were 61-84 years old
- 2 (11%) < 2 years old</p>
- 4 asplenic
- 2 had sickle cell disease (1 asplenic)
- incubation period: 23 384 days
- 5 of 19 (26%) died within days to weeks of diagnosis

- 18 donors implicated
- all IFA positive; only 1 PCR positive
- 12 residents of endemic areas (8 CT, 3 NJ & 1 MA)
- 4 traveled to endemic areas
  - OH to CT, OH to NJ, IN to WI, VA to CT
- 2 implicated in fatal cases
- 1 lost to follow-up & 1 unclear travel history
- none recalled symptoms, only 3 reported tick bite



## **Factors Driving Mitigation Efforts**

- FDA Workshop
- AABB Association Bulletin
- publications
- education
- past failures to act
- babesiosis: nationally notifiable in US
- >100 transmission cases with rising fatalities (n<u>></u>12)





# **Mitigation Strategies**

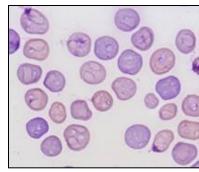
- UDHQ "history of babesiosis"\*
- geographic exclusion\*
- risk-factor questions
- Ieukoreduction
- pathogen reduction
- serologic screening
  - 7 state strategy?
- nucleic acid testing
  - seasonal?

#### \* currently in use

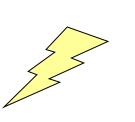


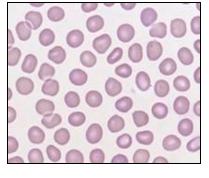
#### **Pathogen Reduction**

- efficacy demonstrated
  - amotosalen + UV light
    - Grellier et al., Transfusion 2008;48:1676-1684.
  - riboflavin + UV light
    - Tonnetti et al., Transfusion 2010;50:1019-1027
- studies limited to apheresis plasma and platelets
- presently, not a viable option in the absence of a whole blood methodology







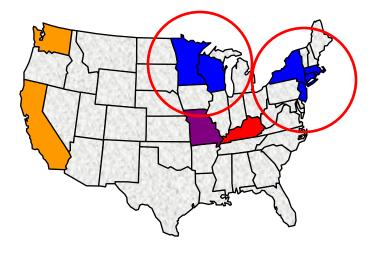


riboflavin + UV



## **Blood Screening Approaches**

- universal screening
- regional testing
- statewide testing
- highly endemic area testing
- CMV model



#### ... if we only had a test!



## **Piloting NAT**

- pilot study of 1,000 CT donations
- collected August/October 2009 from Middlesex and New London Counties
- 1,002 tested to date:
  - 25 (2.5%) IFA positive
  - 3 (0.3%) PCR positive (2 IFA +, 1 IFA -)
    - all identified by first week of September
- 1 apparent window period infection detected
  - number likely low
  - acutely infected donors too sick to donate?
- role for NAT during tick season?



- seasonally triggered
- May through September
- targets acute or "window period" infections
- technologic hurdles remain:
  - PCR sensitivity sufficient, but . . .
  - parasitemia low compared to viral infections
  - requires whole blood
  - limited volume for testing
  - considerations of concentration techniques

